

# Multiparametric imaging of adhesive nanodomains at the surface of *Candida albicans* by atomic force microscopy

Cécile Formosa-Dague, Marion Schiavone, Anita Boisrame, Mathias Richard, Raphael Duval, Etienne Dague

► **To cite this version:**

Cécile Formosa-Dague, Marion Schiavone, Anita Boisrame, Mathias Richard, Raphael Duval, et al.. Multiparametric imaging of adhesive nanodomains at the surface of *Candida albicans* by atomic force microscopy. *Nanomedicine: Nanotechnology, Biology and Medicine*, Elsevier, 2015, 11 (1), pp.57-65. <10.1016/j.nano.2014.07.008>. <hal-01553144>

**HAL Id: hal-01553144**

**<https://hal.laas.fr/hal-01553144>**

Submitted on 7 Sep 2017

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **Multiparametric imaging of adhesive nanodomains at the surface of**  
2 ***Candida albicans* by Atomic Force Microscopy**

3  
4 Cécile Formosa, MS<sup>1,2,3,4</sup>, Marion Schiavone, MS<sup>1,2</sup>, Anita Boisrame, PhD<sup>5,6</sup>, Mathias L. Richard,  
5 PhD<sup>5,6</sup>, Raphaël E. Duval, PhD<sup>3,4,7</sup>, and Etienne Dague, PhD<sup>1,2</sup>

6  
7 <sup>1</sup> CNRS, LAAS, 7 avenue du Colonel Roche, F-31400 Toulouse, France

8 <sup>2</sup> Université de Toulouse ; LAAS, F31400 Toulouse, France

9 <sup>3</sup> CNRS, UMR 7565, SRSMC, Vandœuvre-lès-Nancy, France

10 <sup>4</sup> Université de Lorraine, UMR 7565, Faculté de Pharmacie, Nancy, France

11 <sup>5</sup> INRA, UMR1319 Micalis, F-78352 Jouy-en-Josas, France

12 <sup>6</sup> AgroParisTech, UMR Micalis, F-78850 Thiverval Grignon, France

13 <sup>7</sup> ABC Platform®, Nancy, France

14  
15 Corresponding author: Etienne Dague, LAAS-CNRS, 7 av du Colonel Roche, 31400 Toulouse,  
16 France. Phone number: +33 5 61 33 78 41 Mail : [edague@laas.fr](mailto:edague@laas.fr)

17  
18 Abstract word count: 138 words / Manuscript word count: 4451 words / Number of figures: 5 /  
19 Number of references: 40

20  
21 This work was supported by an ANR young scientist program (AFMYST project ANR-11-JSV5-  
22 001-01 n° SD 30024331) to ED. CF and MS are respectively supported by a grant from  
23 “Direction Générale de l’Armement” (DGA) and from Lallemand SAS.

25 **Abstract**

26 *Candida albicans* is an opportunistic pathogen. It adheres to mammalian cells through a variety  
27 of adhesins that interact with hosts ligands. The spatial organization of these adhesins on the  
28 cellular interface is however poorly understood, mainly because of the lack of an instrument able  
29 to track single molecules on single cells. In this context, the Atomic Force Microscope (AFM)  
30 makes it possible to analyze the force signature of single proteins on single cells. The present  
31 study is dedicated to the mapping of the adhesive properties of *C. albicans* cells. We observed  
32 that the adhesins at the cell surface were organized in nanodomains composed of free or  
33 aggregated mannoproteins. This was demonstrated by the use of functionalized AFM tips and  
34 synthetic amyloid forming/disrupting peptides. This direct visualization of amyloids  
35 nanodomains will help in understanding the virulence factors of *C. albicans*.

36

37

38 **Background**

39           The yeast *Candida albicans* has emerged as a major public health problem these last two  
40 decades. This opportunistic pathogen causes a wide range of infections from surface infections, to  
41 mucosal and blood-stream infections<sup>1</sup>. Whereas mucosal infections are common and occur in  
42 healthy organisms, blood-stream infections are observed only in immunocompromised patients  
43 and are life-threatening. This type of infections, also known as candidaemia, can develop into  
44 disseminated candidiasis when the infection spreads to internal organs, leading to high mortality  
45 rates<sup>2</sup>. In order to colonize and subsequently to disseminate in the blood stream *C. albicans* needs  
46 to adhere to different substrates. This first stage of infection<sup>3</sup> is mediated by adhesins that are  
47 found on the surface of the yeast cell wall. Many of these adhesins are mannoproteins, and  
48 among them, the adhesin family identified as having a major role in host cell attachment is the  
49 Als (Agglutinin-like Sequences) family<sup>4</sup>.

50           The Als were initially reported as having homologies with the proteins responsible for  
51 auto-agglutination in the baker yeast *Saccharomyces cerevisiae*. Eight Als have been identified,  
52 they all are primarily involved in host-pathogen interactions<sup>5</sup>. It was found that there were  
53 amyloid-forming sequences in the Als adhesins of *Candida albicans*<sup>6</sup>. Amyloids are insoluble  
54 fibrillar protein aggregates whose core consists in crystalline arrays of identical sequence in many  
55 molecules of the amyloid protein<sup>7,8</sup>. Cells expressing the Als proteins can rapidly aggregate, and  
56 the aggregation has amyloid-like properties. Like amyloid formation, aggregation ability  
57 propagates through the adherent cell population and depends on conformational changes of the  
58 Als protein. This transition of the conformational state to an aggregative state of the proteins is  
59 characterized by the formation of hydrophobic nanodomains on the entire surface of the cell<sup>9</sup>.

60           A few papers written by Lipke's team were dedicated to the direct visualization of these  
61 nanodomains using fluorescent dyes such as thioflavin T or 8-anilino-1-naphthalene-sulfonic acid

62 (ANS)<sup>6,8,9</sup>. Another technique that can be used to visualize these nanodomains is Atomic Force  
63 Microscopy (AFM). AFM has recently emerged as a valuable tool to study the surface of living  
64 cells<sup>10</sup>, and especially pathogenic cells<sup>11</sup>. This technology has been used by Alsteens *et al.* to  
65 image the formation and propagation of nanodomains in living yeast cells<sup>12</sup> and also to unfold  
66 amyloid proteins from the yeasts surface using Single Molecule Force Spectroscopy<sup>13-15</sup>. To this  
67 end, the authors functionalized AFM tips with antibodies targeted against the Als protein directly  
68 or against an epitope tag present in the Als protein. These studies allowed the authors to localize  
69 the adhesive nanodomains caused by the aggregation of Als proteins at the surface of living yeast  
70 cells, and to unravel the structure of the Als proteins studied by stretching.

71 In our study, we used AFM as an imaging tool to visualize and localize adhesins  
72 nanodomains at the surface of living wild-type *Candida albicans* cells. Using recent  
73 developments in the AFM technology, we have imaged and quantified at the same time the  
74 nanomechanical properties, the adhesiveness (force and nature of the interaction), the size and the  
75 thickness of the nanodomains<sup>16,17</sup>, at high resolution. The data collected showed that these  
76 nanodomains are localized differently at the surface of the cell, depending on the structures  
77 featured by the cells (bud scars, buds). We also showed that there were degrees of adhesiveness,  
78 depending on whether the amyloid proteins had totally aggregated (hydrophobic nanodomains) or  
79 not, and that these degrees of aggregation were directly correlated to the stiffness of the yeast cell  
80 wall. Finally, using force measurements and amyloid forming or inhibiting peptides, we showed  
81 that Als proteins (probably among others) were participating to these nanodomains.

82

## 83 **Methods**

### 84 **Yeasts growth conditions**

85 *Candida albicans* (from ABC Platform® Bugs Bank, Nancy, France) was stocked at -80°C,  
86 revived on Yeast Peptone Dextrose agar (Difco, 242720-500g) and grown in Yeast Peptone  
87 Dextrose broth (Difco, 242820-500g) for 20 hours at 30°C under static conditions.

88

### 89 **Sample preparation for AFM experiments**

90 Yeast cells were concentrated by centrifugation, washed two times in acetate buffer (18 mM  
91 CH<sub>3</sub>COONa, 1 mM CaCl<sub>2</sub>, 1 mM MnCl<sub>2</sub>, pH = 5.2), resuspended in acetate buffer, and  
92 immobilized on polydimethylsiloxane (PDMS) stamps prepared as described by Dague *et al*<sup>18</sup>.  
93 Briefly, freshly oxygen activated microstructured PDMS stamps were covered by a total of 100  
94 µL of the solution of cells and allowed to stand for 15 minutes at room temperature. The cells  
95 were then deposited into the microstructures of the stamp by convective/capillary assembly.  
96 Images were recorded in acetate buffer in Quantitative Imaging™ mode with MLCT AUWH  
97 (Bruker) cantilevers (nominal spring constant of 0.01 N/m). The applied force was kept at 1.5 nN  
98 for imaging and at 0.5 nN for force spectroscopy experiments. The loading rate for imaging was  
99 of 2 500 000 pN/s (acquisition frequency of the force curves is of 25 Hz) and for force  
100 spectroscopy of 75 000 pN/s (acquisition frequency of the force curves is of 1.25 Hz). For  
101 imaging and force spectroscopy, we used an AFM Nanowizard III (JPK Instruments, Berlin,  
102 Germany). The cantilevers spring constants were determined by the thermal noise method<sup>19</sup>. For  
103 all the results presented in this study, silicon nitride AFM tips were bare, except in the case of  
104 figure 4g (lower panel), where a functionalized AFM tip has been used.

105

### 106 **AFM tips fonctionnalization**

107 The functionalized tips were produced according to a french patent of the authors described later  
108 in sensors and actuators<sup>20</sup>. Briefly, AFM tips were functionalized with dendrimers presenting

109 CHO functions able to covalently link with NH<sub>2</sub> functions of proteins. These dendritips were then  
110 incubated with the lectin Concanavalin A (Sigma, L7647-100MG, 100μg/mL) for 1 hour, before  
111 being used for force spectroscopy experiments.

112

### 113 **Results analysis**

114 All results were analyzed using the Data Processing software from JPK Instruments. The stiffness  
115 values measured on cells were determined from the slope of the linear portion of the raw  
116 deflections versus piezo displacement curves, according to:

$$k_{cell} = k \left( \frac{s}{1-s} \right)$$

117 with  $s$  the experimentally accessible slope of the compliance region reached for sufficient loading  
118 forces. In this model, the experimental setup can be represented by two linear springs, one is the  
119 AFM's cantilever, and the other is the cell envelope exhibiting an effective spring constant. It is  
120 then possible to calculate the effective spring constant  $k_{cell}$  of the cell envelope from the  
121 observed slope  $s$  of the force curve and the known spring constant  $k$  of the cantilever<sup>21</sup>.

122

### 123 **Results**

#### 124 *Candida albicans cells display localized adhesiveness*

125 Thanks to our innovative method to immobilize cells into PDMS stamps<sup>18</sup>, and using  
126 Quantitative Imaging™ mode<sup>16</sup>, we were able to image and quantify the adhesive properties of  
127 single *C. albicans* cells at the same time. Figure 1a shows a budding yeast cell; on the  
128 corresponding adhesion image (Figure 1b), we can see that only the bud, and not the mother-cell,  
129 presents adhesives patches. This original result is surprising as non-budding cells are highly  
130 adhesive (see below). This result seems to indicate that the mother-cell cell wall changes during

131 the budding process. As for the cell in figure 1c, this cell displays two bud scars, a common  
132 feature at the surface of yeast cells, which are not adhesive whereas the rest of the cell is. This  
133 type of distribution of the adhesion on yeast cells has already been seen using  
134 immunofluorescence with antibodies targeted against surface proteins of *C. albicans*. Coleman *et*  
135 *al.* for example showed that the Als1 protein was expressed all over *C. albicans* cells, with the  
136 exception of bud scars<sup>22</sup>. The comparison of our results to these data suggests then that the  
137 adhesions probed by AFM might be due to surface proteins, such as Als1 in the case of the cell  
138 presenting bud scars, but perhaps also others adhesins.

139

#### 140 ***C. albicans* cell wall adhesins are able to aggregate into nanodomains**

141 As showed before<sup>9,12</sup>, the proteins expressed at the surface of *C. albicans* cell wall are  
142 able to aggregate, and to form nanodomains. However, these nanodomains have not been yet  
143 characterized at the nanoscale, nor were imaged at high resolution. In fact, these nanodomains  
144 have specific adhesive properties that can be mapped using AFM in the Quantitative Imaging<sup>TM</sup>  
145 mode. High resolution (256 pixels<sup>2</sup>) adhesive images are presented in Figure 2. It shows adhesive  
146 nanodomains, at the surface of a living wild-type *C. albicans* cell. On the cell presented in this  
147 figure (Figure 2a), the corresponding adhesion image shows very distinct adhesive nanodomains  
148 that were probed with bare AFM tips. These nanodomains are homogeneously distributed all over  
149 the cell here, which does not present any morphological features such as buds or bud scars. When  
150 zooming into small areas on top of the cell (white squares on figure 2b), we could measure the  
151 area of each nanodomain. On this cell and on another one showed in Figure 3a, 60 nanodomains  
152 areas were measured; the values obtained plotted on figure 2e shows that nanodomains have an  
153 average area of  $0.09 \pm 0.03 \mu\text{m}^2$ . This corresponds to an average diameter of 170 nm, which  
154 confirms the nanoscale of these nanodomains. Some of the nanodomains are also higher than the



155 rest of the cell wall. When the whole cell is imaged, it is not visible; however, specific analysis of  
156 the Figure 3d and the graphic 3h representing the topography of the cell surface revealed  
157 nanodomains that had a different height compared to the rest of the cell. The cross-sections taken  
158 along the blue line showed the height of a nanodomain of 20 nm. Once again, this confirmed the  
159 nanoscale of the nanodomains at the surface of *C. albicans*.

160

### 161 ***Different nanodomains have different nanomechanical properties***

162 Adhesion is measured as the rupture force recorded when retracting the tip from the surface,  
163 when approaching and pulling with the tip on the cell wall, thus AFM makes it possible to  
164 measure nanomechanical properties of living cells. Here we choose to use an analysis based on  
165 the Hooke model which considers the coupled cantilever / cell wall as a spring. The stiffness  
166 values measured on cells were determined from the slope of the linear portion of the raw  
167 deflections versus piezo displacement curves, according to:

$$k_{cell} = k \left( \frac{s}{1-s} \right)$$

168 with  $s$  the experimentally accessible slope of the compliance region reached for sufficient loading  
169 forces. Indeed, the most interesting result in this study is the correlation that can be directly made  
170 between the adhesiveness of the nanodomain, and its stiffness. Nanodomains on figure 3e  
171 (adhesion map) circled in red were found on the stiffness image (figure 3f) circled in black; they  
172 correspond to the zones where the stiffness of the cell wall is increased, to  $13.4 \pm 0.3$  nN/ $\mu$ m. As  
173 for less adhesive nanodomains, they do not present any difference in stiffness from the rest of the  
174 cell, and are  $12.4 \pm 0.2$  nN/ $\mu$ m. The 3D-view of the adhesion, mapped with the stiffness (figure  
175 3g) illustrates this clear correlation; the more adhesive the nanodomain is, the stiffer it is.

176 Another fascinating point is that for the more adhesive nanodomains, the retract force  
177 curves present typical hydrophobic adhesions<sup>23-25</sup>, with adhesions occurring immediately when  
178 the tips is retracted from the surface. Force curves from the other, less adhesive, nanodomains  
179 presented retract adhesions resembling to proteins unfolding, occurring several nanometers after  
180 the tip withdrawal. Therefore it seems that the nanodomains are of 2 different natures. There is a  
181 class of nanodomains, hydrophobic, higher and stiff, and another class displaying proteins  
182 unfolding properties, as soft as the rest of the cell wall. What is the molecular nature of these 2  
183 types of nanodomains, and are they correlated?

184

#### 185 *Understanding the adhesive properties of the 2 nanodomains classes*

186 To answer the previous question, we monitored the retract force curves recorded on the  
187 nanodomains (Figure 4). We found the same correlation as in figure 3; the force curves recorded  
188 on an adhesive nanodomains presented hydrophobic retract adhesions, whereas the force curves  
189 recorded on a less adhesive nanodomain presented protein, unfolding like, profiles. In order to  
190 determine the nature of these last unfoldings, we probed the surface of *C. albicans* cells with an  
191 AFM tip functionalized with Concanavalin A (ConA), a protein that interacts with yeast  
192 mannoproteins, such as surface adhesins. The resulting force curves (figure 4g) showed retract  
193 adhesions displaying unfoldings of different lengths, but with a similar profile. We also observed  
194 condensed spikes with adhesion forces between 0 and 50 pN. This value was consistent with  
195 specific interactions between ConA and mannoproteins<sup>26</sup>.

196 In a previous study conducted in 2009<sup>14</sup> by Alsteens *et al.*, adhesins (Als5) were unfolded  
197 from the surface of live *S. cerevisiae* cells overexpressing this protein. The retract force curves  
198 obtained in this study show high similarity with the ones we obtain here with functionalized AFM  
199 tips, with the presence of serin-threonin rich segments (condensed pikes on figure 4g). We can

200 therefore, based on this comparison with the data of the literature, conclude that the less adhesive  
201 nanodomains at the surface of live *C. albicans* are composed of free adhesins, and maybe of Als  
202 proteins. However, since all adhesins (like Als, Hwp1, Eap1, Rbt1 etc) are mannoproteins, we  
203 cannot, at this stage, make a statement on which adhesins are unfolded here.

204 As for the second type of nanodomains, the hydrophobic ones, our hypothesis is that they  
205 are composed of the same proteins as the less adhesive ones. In fact, adhesins (like Als) display  
206 amyloids sequences located on a domain of the protein called T, that enable them to change their  
207 conformation<sup>6</sup> and to aggregate into amyloid nanodomains. And when this phenomena is started,  
208 it propagates to the whole cell<sup>9</sup>. We therefore made the hypothesis that the adhesive nanodomains  
209 are in fact amyloid nanodomains, made of Als proteins.

210

### 211 ***From adhesins to amyloid nanodomains: the role of Als proteins***

212 To verify this hypothesis, and according to the literature on Als proteins, we synthesized a  
213 peptide exhibiting the same sequence as the one of the T domain of the Als1/3/5 proteins. We  
214 then put this peptide in the presence of the cells in order to trigger the amyloids formation. We  
215 also synthesized the same peptide, but with a mutation on one amino acid (V326N peptide), in  
216 order to obtain a peptide that inhibits the formation of the amyloid nanodomains<sup>8</sup>. Since Als3 is  
217 only expressed on the surface of hyphae, we will only be able to generate or destroy the amyloid  
218 formation of Als 1 and 5. The results presented in figure 5 showed a cell before and after adding  
219 the mutated peptide. We can clearly see on these adhesion images the loss of general adhesion  
220 and of two nanodomains at the center of the cell. It seems like the mutated peptide disrupted the  
221 amyloids at the surface of the cell. And the other way around, when cells were incubated with the  
222 amyloid forming peptide, we could observe the formation of the nanodomains at the surface of  
223 the cells, as it is showed on the adhesion images on local areas on top of *C. albicans* cells in

224 figure 5c, d and e. These results allow us to conclude that the proteins at the origin of the  
225 nanodomains are then mannoproteins, more specifically Als1 or 5, or both, that form amyloids.

226

## 227 **Discussion**

228 We show in this study that wild-type live *C. albicans* cells exhibit extraordinary adhesive  
229 properties. In the case of budding cells, placed in acetate buffer at 25°C for 2h, we observed that  
230 the mother cell is not adhesive and that only the bud presents adhesive nanodomains. On the  
231 contrary, we show that non budding cells are covered by adhesive nanodomains, in the same  
232 experimental conditions. This illustrates the amazing plasticity of this species<sup>27,28</sup> able to grow as  
233 a commensal or as a pathogen<sup>29,30</sup>, in all the parts of the intestinal track, but also on the vaginal  
234 mucosa, as unicellular budding cells or as filamentous hyphae. Moreover its cell wall is  
235 permanently remodeled as a reaction to its environment (temperature, pH, dissolved O<sub>2</sub>, ions,  
236 interacting surface/cells/bacteria) what makes it challenging to reproduce the experimental  
237 conditions inducing a certain cell wall phenotype. We then demonstrate that the molecules at the  
238 origin of these adhesions could aggregate into nanodomains, which can be probed at high  
239 resolution using a suited AFM mode, QI<sup>TM</sup>. These nanodomains are different in terms of level of  
240 adhesiveness, which is a property directly correlated to their stiffness and to the hydrophobic  
241 state or not of the molecule at the origin of these nanodomains. We then went further in the study,  
242 using functionalized AFM tips, and were able to determine that the less adhesive nanodomains  
243 were formed by mannoproteins that can interact specifically with Concanavalin A. These  
244 mannoproteins are able to aggregate to form the adhesive nanodomains because they have  
245 amyloid properties as we showed in figure5.

246 Amyloid aggregation is a primitive<sup>31</sup> and very stable<sup>32</sup> protein folding and a common  
247 structural motif. It is a cross  $\beta$ -sheet quaternary structure that usually auto-aggregates as fibrils. It

248 has been, first, associated with neurodegenerative diseases like Alzheimer, Parkinson, or  
249 Creutzfeldt-Jakob diseases. However it is more and more unclear if the amyloids lesions are the  
250 cause or a consequence of the disease. Amyloid aggregates are now described as functional  
251 proteins assembly and can be found from bacteria to humans<sup>33</sup>. In microorganisms, amyloid has  
252 been described as a functional coat<sup>34</sup>. It consists in curli (*E. coli*), chaperons (*Streptomyces*) or  
253 hydrophobins (*Aspergillus etc*); all of these proteins are implicated in adhesion to the host and in  
254 the invasion, infection process. It is now well known that adhesins (and especially *AIs*) of *C.*  
255 *albicans* have amyloid-forming sequences<sup>6,35</sup> and that these proteins form domains involved in  
256 cell aggregation or biofilm formation<sup>36</sup>. Nevertheless the characterization, structure and  
257 properties of the amyloids adhesive nanodomains remain unclear.

258 In this work we measured for the first time the nanoscale size of amyloid domains  
259 (average area of  $0.09 \mu\text{m}^2$ ) at the surface of live *C. albicans* cells. The domains are of 2 different  
260 classes. Some present the characteristic of individual proteins whereas the others are  
261 hydrophobic, stiffer than the rest of the cell ( $13.4 \pm 0.3 \text{ nN}/\mu\text{m}$  compared to  $12.4 \pm 0.2 \text{ nN}/\mu\text{m}$ ),  
262 and are slightly protruding. It means that there is a state modification from soluble proteins into  
263 insoluble proteins, which is a characteristic of amyloid structures. This transformation is  
264 dependent on the proteins concentration and can only occur when the protein density exceeds a  
265 threshold. The roles of the two classes of domains are probably different. On one hand we could  
266 hypothesize that the hydrophobic nanodomains were involved in the cell adhesion to abiotic  
267 hydrophobic surfaces or to cell membrane as it is known that membrane binding is an inherent  
268 property of amyloid aggregates<sup>37,38</sup>. Amyloid aggregation is also a way to store proteins, in a  
269 limited space and to sort them when required. This has been demonstrated for hormones in  
270 secretory granules<sup>39,40</sup>. Thus *C. albicans* may store some adhesins for the subsequent invasion  
271 phases. On the other hand the protein like domains may be responsible for specific adhesion to

272 fibronectin and other extra cellular proteins of the matrix. It seems rational that several adhesins,  
273 brought together, would be more efficient in a binding process than a single adhesion. This  
274 finding has to be added to *C. albicans* plasticity<sup>28</sup> and participate to explain its remarkable  
275 adaptation and pathogenicity.

276           However, there are still many things to explore on the cell wall of *C. albicans*, and future  
277 work will be dedicated to exploring the changes appearing on the mother cell during the budding  
278 process.

279

## 280 **References**

281

- 282 1. Gow NA, Hube B. Importance of the *Candida albicans* cell wall during commensalism and  
283 infection. *Curr Opin Microbiol.* 2012;**15**:406–412.
- 284 2. Poulain D. *Candida albicans* , plasticity and pathogenesis. *Crit Rev Microbiol.* 2013:1–10.
- 285 3. Naglik JR, Moyes DL, Wächtler B, Hube B. *Candida albicans* interactions with epithelial  
286 cells and mucosal immunity. *Microbes Infect.* 2011;**13**:963–976.
- 287 4. Hoyer LL. The ALS gene family of *Candida albicans*. *Trends Microbiol.* 2001;**9**:176–180.
- 288 5. Hoyer LL, Green CB, Oh S-H, Zhao X. Discovering the secrets of the *Candida albicans*  
289 agglutinin-like sequence (ALS) gene family – a sticky pursuit. *Med Mycol.* 2008;**46**:1–15.
- 290 6. Ramsook CB, Tan C, Garcia MC, Fung R, Soybelman G, Henry R, et al. Yeast Cell  
291 Adhesion Molecules Have Functional Amyloid-Forming Sequences. *Eukaryot Cell.*  
292 2010;**9**:393–404.
- 293 7. Sawaya MR, Sambashivan S, Nelson R, Ivanova MI, Sievers SA, Apostol MI, et al. Atomic  
294 structures of amyloid cross- $\beta$  spines reveal varied steric zippers. *Nature.* 2007;**447**:453–457.
- 295 8. Garcia MC, Lee JT, Ramsook CB, Alsteens D, Dufrêne YF, Lipke PN. A Role for Amyloid  
296 in Cell Aggregation and Biofilm Formation. *PLoS ONE.* 2011;**6**:e17632.
- 297 9. Rauceo JM, Gaur NK, Lee K-G, Edwards JE, Klotz SA, Lipke PN. Global Cell Surface  
298 Conformational Shift Mediated by a *Candida albicans* Adhesin. *Infect Immun.*  
299 2004;**72**:4948–4955.

- 300 10. Müller DJ, Dufrêne YF. Atomic force microscopy: a nanoscopic window on the cell  
301 surface. *Trends Cell Biol.* 2011;**21**:461–469.
- 302 11. Alsteens D, Beaussart A, El-Kirat-Chatel S, Sullan RMA, Dufrêne YF. Atomic Force  
303 Microscopy: A New Look at Pathogens. *PLoS Pathog.* 2013;**9**:e1003516.
- 304 12. Alsteens D, Garcia MC, Lipke PN, Dufrêne YF. Force-induced formation and propagation  
305 of adhesion nanodomains in living fungal cells. *Proc Natl Acad Sci.* 2010;**107**:20744 –  
306 20749.
- 307 13. Beaussart A, Alsteens D, El-Kirat-Chatel S, Lipke PN, Kucharíková S, Van Dijck P, et al.  
308 Single-Molecule Imaging and Functional Analysis of Als Adhesins and Mannans during  
309 *Candida albicans* Morphogenesis. *ACS Nano.* 2012;**6**:10950–10964.
- 310 14. Alsteens D, Dupres V, Klotz SA, Gaur NK, Lipke PN, Dufrêne YF. Unfolding Individual  
311 Als5p Adhesion Proteins on Live Cells. *ACS Nano.* 2009;**3**:1677–1682.
- 312 15. Alsteens D, Ramsook CB, Lipke PN, Dufrêne YF. Unzipping a Functional Microbial  
313 Amyloid. *ACS Nano.* 2012;**6**:7703–7711.
- 314 16. Chopinet L, Formosa C, Rols MP, Duval RE, Dague E. Imaging living cells surface and  
315 quantifying its properties at high resolution using AFM in QI<sup>TM</sup> mode. *Micron.* 2013;**48**:26–  
316 33.
- 317 17. Dufrêne YF, Martínez-Martín D, Medalsy I, Alsteens D, Müller DJ. Multiparametric  
318 imaging of biological systems by force-distance curve-based AFM. *Nat Methods.*  
319 2013;**10**:847–854.
- 320 18. Dague E, Jauvert E, Laplatine L, Viallet B, Thibault C, Ressler L. Assembly of live micro-  
321 organisms on microstructured PDMS stamps by convective/capillary deposition for AFM  
322 bio-experiments. *Nanotechnology.* 2011;**22**.
- 323 19. Hutter JL, Bechhoefer J. Calibration of atomic-force microscope tips. *Rev Sci Instrum.*  
324 1993;**64**:1868–1873.
- 325 20. Jauvert E, Dague E, Séverac M, Ressler L, Caminade A-M, Majoral J-P, et al. Probing  
326 single molecule interactions by AFM using bio-functionalized dendritips. *Sens Actuators B*  
327 *Chem.* 2012;**168**:436–441.
- 328 21. Arnoldi M, Fritz M, Bäuerlein E, Radmacher M, Sackmann M, Boulbitch A. Bacterial  
329 turgor pressure can be measured by atomic force microscopy. *Phys Rev E.* 1999;**62**:1034-1044  
330
- 331 22. Coleman DA, Oh S-H, Zhao X, Hoyer LL. Heterogeneous distribution of *Candida albicans*  
332 cell-surface antigens demonstrated with an Als1-specific monoclonal antibody.  
333 *Microbiology.* 2010;**156**:3645–3659.
- 334 23. Dague E, Alsteens D, Latge JP, Verbelen C, Raze D, Baulard AR, et al. Chemical force  
335 microscopy of single live cells. *Nano Lett.* 2007;**7**:3026–3030.

- 336 24. Alsteens D, Dague E, Rouxhet PG, Baulard AR, Dufrene YF. Direct measurement of  
337 hydrophobic forces on cell surfaces using AFM. *Langmuir*. 2007;**23**:11977–11979.
- 338 25. Dague E, Alsteens D, Latgé J-P, Dufrêne YF. High-Resolution Cell Surface Dynamics of  
339 Germinating *Aspergillus fumigatus* Conidia. *Biophys J*. 2008;**94**:656–660.
- 340 26. Alsteens D, Dupres V, Evoy KM, Wildling L, Gruber HJ, Dufrêne YF. Structure, cell wall  
341 elasticity and polysaccharide properties of living yeast cells, as probed by AFM.  
342 *Nanotechnology*. 2008;**19**.
- 343 27. Odds F. *Candida* and candidosis. A review and bibliography. 2nd edition. London: Bailliere  
344 Tindall; 1988.
- 345 28. Poulain D. *Candida albicans*, plasticity and pathogenesis. *Crit Rev Microbiol*. 2013:1–10.
- 346 29. Gow NA, Hube B. Importance of the *Candida albicans* cell wall during commensalism and  
347 infection. *Curr Opin Microbiol*. 2012;**15**:406–412.
- 348 30. Gow NAR, van de Veerdonk FL, Brown AJP, Netea MG. *Candida albicans* morphogenesis  
349 and host defence: discriminating invasion from colonization. *Nat Rev Micro*. 2012;**10**:112–  
350 122.
- 351 31. Greenwald J, Riek R. Biology of Amyloid: Structure, Function, and Regulation. *Structure*.  
352 2010;**18**:1244–1260.
- 353 32. Perczel A, Hudáky P, Pálfi VK. Dead-End Street of Protein Folding: Thermodynamic  
354 Rationale of Amyloid Fibril Formation. *J Am Chem Soc*.; **129**:14959–14965.
- 355 33. Fowler DM, Koulov AV, Balch WE, Kelly JW. Functional amyloid--from bacteria to  
356 humans. *Trends Biochem Sci*. 2007;**32**:217–224.
- 357 34. Gebbink MFBG, Claessen D, Bouma B, Dijkhuizen L, Wösten HAB. Amyloids — a  
358 functional coat for microorganisms. *Nat Rev Microbiol*. 2005;**3**:333–341.
- 359 35. Otoo HN, Lee KG, Qiu W, Lipke PN. *Candida albicans* Als Adhesins Have Conserved  
360 Amyloid-Forming Sequences. *Eukaryot Cell*. 2008;**7**:776–782.
- 361 36. Garcia MC, Lee JT, Ramsook CB, Alsteens D, Dufrêne YF, Lipke PN. A Role for Amyloid  
362 in Cell Aggregation and Biofilm Formation. *PLoS ONE*. 2011;**6**:e17632.
- 363 37. Sparr E, Engel MFM, Sakharov DV, Sprong M, Jacobs J, de Kruijff B, et al. Islet amyloid  
364 polypeptide-induced membrane leakage involves uptake of lipids by forming amyloid  
365 fibers. *FEBS Lett*. 2004;**577**:117–120.
- 366 38. Gellermann GP, Appel TR, Tannert A, Radestock A, Hortschansky P, Schroeckh V, et al.  
367 Raft lipids as common components of human extracellular amyloid fibrils. *Proc Natl Acad*  
368 *Sci U S A*. 2005;**102**:6297–6302.



369 39. Maji SK, Perrin MH, Sawaya MR, Jessberger S, Vadodaria K, Rissman RA, et al.  
370 Functional Amyloids As Natural Storage of Peptide Hormones in Pituitary Secretory  
371 Granules. *Science*. 2009;**325**:328–332.

372 40. Dannies PS. Concentrating hormones into secretory granules: layers of control. *Mol Cell*  
373 *Endocrinol*. 2001;**177**:87–93.

374

### 375 **Figure Caption**

376 **Figure 1. Localization of the adhesive properties of *C. albicans* cells.** (a) Height image (z-  
377 range = 1.5  $\mu\text{m}$ ) of a budding *C. albicans* cell in a PDMS stamp, and (b) adhesion image  
378 corresponding to the height image. On (a), MC stands for Mother cell, BC stands for Budding  
379 cell, and the red dotted line represents the demarcation between the two different cells. (c) Height  
380 image (z-range = 3.5  $\mu\text{m}$ ) of a single *C. albicans* cell exhibiting two bud scars, and (d) adhesion  
381 image corresponding to the height image.

382

383 **Figure 2. Imaging of the adhesive domains of *C. albicans* cells in acetate buffer, at 25°C for**  
384 **2 hours.** (a) Height image (z-range = 2.5  $\mu\text{m}$ ) of a single *C. albicans* cell in a  
385 polydimethylsiloxane (PDMS) stamp, and (b) adhesion images corresponding to the height  
386 images. (c, d) Adhesion images of small areas on top of the cell, represented by the white squares  
387 in b. (e) distribution of the areas values of the domains in c and d.

388

389 **Figure 3. Nanomechanics of the adhesive domains of *C. albicans* cells.** (a) Height image (z-  
390 range = 2.5  $\mu\text{m}$ ) of a *C. albicans* cell in a PDMS stamp, (b) corresponding adhesion image, and  
391 (c) corresponding stiffness image. (d) Height image (z-range = 100 nm) of a small area on top of  
392 the cell, represented by the white square on (a), (e) corresponding adhesion image and (f)  
393 corresponding stiffness image. Note that the adhesive nanodomains circled in red on (e) are also

394 found on the stiffness image (black circles on f). (g) is a 3D-image of the adhesion mapped with  
395 the stiffness. (h) cross-section taken along the blue line on (d), and (i), distribution of the stiffness  
396 values corresponding to the yeast cell wall and the less adhesive domains (blue columns) or to the  
397 most adhesive domains (yellow columns).

398  
399 **Figure 4. Adhesion force curves of *C. albicans* adhesive domains.** (a) Height image (z-range =  
400 4.0  $\mu\text{m}$ ) of a *C. albicans* cell in a PDMS stamp. (b) Adhesion image of a small area on top of the  
401 cell, represented by the white square on (a). (c and d) representative force curves obtained on the  
402 zones indicated by the arrows on (c). (e) Height image (z-range = 2.5  $\mu\text{m}$ ) of a *C. albicans* cell in  
403 a PDMS stamp and (f) corresponding adhesion image recorded with a bare tip. (g) representative  
404 force curves obtained in the zone delimited by a white square on (f) with the Con A tip.

405  
406 **Figure 5. Imaging of the adhesive domains of *C. albicans* cells treated with Als1, 3, 5p**  
407 **amyloid disrupting peptide (V326N peptide) or Als1, 3, 5p amyloid forming peptide (Als**  
408 **peptide).** (a) Adhesion image of a single *C. albicans* cell in a PDMS stamp and (b)  
409 corresponding adhesion image after adding V326N peptide. (c, d and e) Adhesion images of  
410 small areas on top of a *C. albicans* cell after adding the Als peptide.

411  
412